SOLIS BIODYNE

FIREPol® DNA Polymerase

(5 U/µl)

Catalogue Number	Pack Size (5 U/µl)
01-01-0000S	100 U
01-01-00500	500 U
01-01-01000	1000 U
01-01-02000	2000 U

Shipping:

At room temperature

Batch Number and Expiry Date:

See vial

Stability at room temperature 1 MONTH

Store at -20 °C

Storage and Stability:

- Routine storage at - 20 °C until expiry date
- Can be stored at +4 °C for up to 6 months
- Stability at room temperature (15–25 °C) for 1 month
- Freeze-thaw stability: 30 cycles

Reaction setup:

At room temperature

Manufactured by Solis BioDyne, in compliance with the ISO 9001 and ISO 13485 certified Quality Management System.

Product description:

- FIREPol® is a highly processive, thermostable DNA Polymerase. Due to its genetic modifications, it has an enhanced stability at room temperature with no activity loss for up to 1 month.
- Recommended for routine applications (fragment up to 3 kb from genomic DNA).
- Possesses 5′→3′ polymerase and 5′→3′ endonuclease activity, as well as a non-template-dependent terminal transferase activity, but lacks a 3′→5′ exonuclease (proofreading) activity making the generated product suitable for TA-cloning.
- The fidelity of FIREPol® is similar to a regular Taq DNA Polymerase (error rate per nucleotide ca 2.5 x10^{-5}).

Contents:

Component	Catalogue Number 01-01-0000S	01-01-00500	01-01-01000	01-01-02000
FIREPol® DNA Polymerase (5 U/µl)	100 U / 20 µl	500 U / 100 µl	1000 U / 200 µl	2000 U / 400 µl
FIREPol® 10x Buffer B	500 µl	2.5 ml	5.0 ml	2 x 5.0 ml
FIREPol® 10x Buffer BD	500 µl	2.5 ml	5.0 ml	2 x 5.0 ml
25 mM $MgCl_2$	500 µl	2.5 ml	5.0 ml	2 x 5.0 ml
10x Solution S	100 µl	100 µl	500 µl	500 µl

- **FIREPol® DNA Polymerase** (5 units/µl) in 20 mM Tris-HCl pH 8.7 at 25 °C, 100 mM KCl, 0.1 mM EDTA, 50% glycerol (v/v), and stabilizers.
- **FIREPol® 10x Buffer B** (Mg^{2+} free, with detergent): 0.8 M Tris-HCl, 0.2 M $(NH_4)_2SO_4$, 0.2% w/v Tween-20.

  FIREPol® 10x Buffer B contains non-ionic detergent suppressing inhibitory effects of the traces of DNA extraction buffer and enhancing PCR yield and efficiency.

- **FIREPol® 10x Buffer BD** (Mg^{2+} and detergent free): 0.8 M Tris-HCl, 0.2 M $(NH_4)_2SO_4$.
- **25 mM $MgCl_2$**
- **10x Solution S** is an additive that facilitates amplification of difficult templates (e.g. GC-rich DNA templates).

  This solution should be used at a defined final concentration (1x, 2x or 3x solution). 10x Solution S is NOT a reaction buffer and should be used ONLY IF non-specific amplification occurs.

Additional reagents required:

- Template DNA
- Gene-specific primer pair
- dNTP Mix (20 mM of each, Cat. No. 02-31-00020)
- Nuclease-free PCR Grade Water (Cat. No. water-025)

Step-by-step guidelines:

1. Thaw the reagents at room temperature. Mix each reagent by gentle vortexing or pipetting up and down, then centrifuge briefly.
2. Prepare a reaction mix at room temperature. Add all required components except the template DNA.

Component	Volume[1]	Final conc.
FIREPol® DNA Polymerase (5 U/µl)	0.08–0.2 µl	0.02–0.05 U/µl
FIREPol® 10x Buffer B or BD	2 µl	1x
25 mM $MgCl_2$	1.2–2 µl	1.5–2.5 mM
dNTP Mix (20 mM of each)	0.2 µl	200 µM of each
Forward Primer (10 µM)	0.2–0.6 µl	100–300 nM
Reverse Primer (10 µM)	0.2–0.6 µl	100–300 nM
10x Solution S (optional)	2, 4 or 6 µl	1x, 2x or 3x
Template DNA (added at step 4)	Variable	Variable[2]
Nuclease-free water	up to 20 µl	
Total reaction volume	**20 µl**	

[1] Scale all components proportionally according to sample number and reaction volumes. Make sure you use enough of each reagent for your reactions, plus 10% extra volume to accommodate pipetting errors.

[2] For low complexity templates (i.e. plasmid, lambda), use 20 pg–2 ng of DNA per 20 µl reaction. For higher complexity templates (i.e. gDNA), use 2 ng–200 ng of DNA per 20 µl reaction.

3. Mix the reaction mix thoroughly, then centrifuge briefly. Dispense appropriate volumes of mix into PCR wells.

4. Add template DNA to the PCR wells. Seal the wells using the procedure recommended for the cycling instrument being used, and centrifuge the reactions briefly.

5. Incubate your PCR reactions in thermal cycler as follows.

Step	Temperature	Time	Cycles
Initial denaturation[1]	95 °C	3–5 min	1
Denaturation	95 °C	15–30 sec	26–35
Annealing[2]	50–68 °C	30–60 sec	
Extension[2]	72 °C	45 sec–4 min	
Final extension	72 °C	5–10 min	1

[1] Complex templates, such as gDNA, require longer time to denature (5 min). With low complexity templates (i.e. lambda, plasmid DNA), initial denaturation time can be reduced to 3 min.

[2] The annealing temperature depends on the melting temperature of the primers. Extension time depends on the length of the fragment to be amplified. A time of 1 min/kb is recommended.

Recommendations for a successful PCR experiment

Prerequisites for a successful PCR include the design of optimal primers, the use of high-quality template DNA and appropriate concentrations of reaction components.

Use dedicated software, such as Primer3 and NCBI Primer-BLAST to design target-specific primers. The optimal primer length is 20–30 bp, with GC-content 35–65% and calculated melting

temperatures (T_m) 60–70 °C. T_m of the two primers should not differ by more than 3 °C. Analyze your primers for self-complementarity and stable secondary structures, presence of secondary structures increases probability of mis-priming and primer-dimers formation.

The integrity, purity and concentration of the DNA template should be suitable for the PCR experiment. Always include a no-template control (NTC) by replacing the DNA template with the same volume of nuclease-free water.

Please see the Troubleshooting Guide below for suggestions and help with specific problems.

Troubleshooting Guide

No or low PCR yield

- Cycling conditions are not optimal – decrease the primer annealing temperature (T_a); if needed determine the optimal T_a by running a temperature gradient; increase the extension time (if amplifying a long target); increase the number of cycles by 3–5.
- Poor quality of template – check the template's purity and integrity, ensure that your template doesn't contain PCR inhibitors.
- Template concentration is too low – increase the concentration of DNA template.
- Primer concentration is not optimal – titrate primer concentration (final concentration 100–300 nM of each); ensure that both primers have the same concentration.
- Reaction components are degraded – check the storage conditions and expiry date of the reagents; perform a positive

control with template DNA and/or reagents previously known to amplify.

Non-specific products

- Non-specific amplification – use hot-start PCR enzyme (e.g. HOT FIREPol DNA Polymerase, Cat. No. 01-02-00500); ensure that your primers are target-specific.
- Primer concentration is not optimal – titrate primer concentration (final concentration 100–300 nM of each); too high primer concentration can reduce the binding specificity, resulting in unwanted products.
- Primer annealing temperature (T_a) is too low – increase the T_a; keep your primer annealing temperature 2–5 °C below the T_m of the primer having the lowest T_m.
- Too many cycles – reduce the cycle number by 3–5.
- Contamination – to avoid contamination, work in dedicated space, keep pre- and post-amplification areas separate, decontaminate your surfaces and equipment, if possible, aliquot your reagents into smaller volumes to prevent contamination of stock solutions.

Smearing in electrophoresis

- Too much template – load lower amount or prepare serial dilutions of template.
- Too many cycles – reduce the cycle number by 3–5.
- Extension time is too long – reduce extension time.
- Primer design is not optimal – review your primers and redesign the primers if needed.
- Enzyme concentration is too high – decrease the amount of enzyme in final solution by 0.005 U/µl increments (optimal polymerase concentration in final PCR solution is 0.02–0.05 U/µl).

Source:

E. coli strain that carries an overproducing plasmid with a modified gene of *Thermus aquaticus* DNA Polymerase.

Unit definition:

One unit is defined as the amount of enzyme required to catalyze the incorporation of 10 nmol of dNTPs into an acid-insoluble form in 30 minutes at 74 °C.

Quality control:

The enzyme is free of nicking and priming activities, exonucleases and non-specific endonucleases. SDS/PAGE 95 kD band, >98% pure. Activity and stability tested via thermocycling. The error rate per nucleotide per cycle is ca 2.5 $x10^{-5}$; the accuracy is ca 4 $x10^{4}$. Estimated half-life at 95 °C is 1.5 hours.

Safety precautions:

Please refer to Safety Data Sheet for more information.

Technical support:

Contact your sales representative for any questions or send an email to support@solisbiodyne.com

Revised 04.02.2020

SOLIS BIODYNE